

# Amplification and overexpression of a novel m6A reader FXR1 are associated with poor prognosis in head and neck squamous cell carcinoma

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A – research concept and design; B – collection and/or assembly of data; C – data analysis and interpretation;

D – writing the article; E – critical revision of the article; F – final approval of the article

Dental and Medical Problems, ISSN 1644-387X (print), ISSN 2300-9020 (online)

Dent Med Probl. 2026;63(2):397–405

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## Funding sources

None declared

## Conflict of interest

None declared

## Acknowledgements

None declared

Received on January 8, 2024

Reviewed on February 26, 2024

Accepted on March 2, 2024

Published online on April 22, 2026

## Cite as

Manicka Vasagam J, Kannan B, Jayaseelan VP, Arumugam P. Amplification and overexpression of a novel m6A reader FXR1 are associated with poor prognosis in head and neck squamous cell carcinoma. *Dent Med Probl.* 2026;63(2):397–405. doi:10.17219/dmp/185583

## DOI

10.17219/dmp/185583

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## Abstract

**Background.** Fragile X-related protein 1 (FXR1), an RNA-binding protein (RBP) implicated in various cancers, has emerged as a potential contributor to the progression of head and neck squamous cell carcinoma (HNSCC).

**Objectives.** The aim of the study was to comprehensively analyze FXR1 genetic alterations, mRNA expression, protein levels, and immune infiltration in HNSCC, in order to elucidate its molecular role and therapeutic potential.

**Material and methods.** Data from The Cancer Genome Atlas (TCGA)—HNSCC cohort was analyzed. Copy number variation (CNV) was assessed using cBioPortal, FXR1 expression profiling was performed using the University of Alabama at Birmingham CANcer data analysis Portal (UALCAN), and immunohistochemistry (IHC) examination was conducted using the Human Protein Atlas (HPA). The TIMER 2.0 and TISIDB databases were employed for immune infiltration analysis.

**Results.** Genetic alterations in *FXR1*, including DNA amplification, were observed in 22% of HNSCC patients and were significantly correlated with elevated mRNA expression. FXR1 mRNA and protein levels were upregulated in HNSCC tumors, and this upregulation was associated with poor prognosis. The expression of *FXR1* was also correlated with clinical features, including disease stage, grade, metastasis, and human papillomavirus (HPV) status. Immune infiltration analysis revealed a negative correlation between *FXR1* mRNA expression and CD8<sup>+</sup> T cells, suggesting a potential role in immune modulation.

**Conclusions.** FXR1 may serve as a potential biomarker and immune regulator in HNSCC, with genetic alterations influencing its mRNA expression. This study sheds light on the multifaceted role of FXR1 in the progression of HNSCC, and suggests potential avenues for targeted therapeutic strategies. Further research, including functional experiments and large-scale cohort analyses, is warranted to validate and translate these findings into clinical applications.

**Keywords:** TCGA, prognosis, copy number variation, head and neck carcinoma, FXR1

## Highlights

- FXR1 may serve as a potential biomarker and immune regulator in head and neck squamous cell carcinoma (HNSCC), with genetic alterations influencing its mRNA expression.
- FXR1 appears to play a multifaceted role in HNSCC progression, including potential involvement in tumor–immune interactions, and may represent a target for future therapeutic strategies.
- Further research, including functional studies and large-scale cohort analyses, is required to validate these findings and support their clinical translation.

## Introduction

Head and neck squamous cell carcinoma (HNSCC) constitutes a diverse group of tumors, predominantly squamous cell carcinomas (SCCs), accounting for approx. 90% of diagnoses and ranking as the 6<sup>th</sup> most prevalent cancer worldwide.<sup>1,2</sup> Oropharyngeal cancers are primarily driven by human papillomavirus (HPV), while alcohol consumption and tobacco use are risk factors for cancers of the oral cavity, hypopharynx and larynx. Notably, HPV-positive HNSCC generally exhibits a more favorable prognosis.<sup>3</sup> The development of HNSCC involves intricate genetic and epigenetic alterations that contribute to hallmark features of cancer, including uncontrolled proliferation and metastasis.<sup>4–8</sup>

The discovery of human fragile X-related protein 1 (FXR1) (3q28) resulted from fragile X messenger ribonucleoprotein 1 (FMR1) sequencing, revealing an autosomal gene with 60% amino acid sequence identity to other family members.<sup>9</sup> FXR1 shares a common structure but differs in its C-termini, implying distinct roles. The protein contains several domains, including a Tandem agent-like domain, a non-classical nuclear localization signal (NLS), 3 K homology (KH) domains, a nuclear export signal (NES), and an arginine–glycine–glycine (RGG) box, classifying FXR1 as an RNA-binding protein (RBP). Exportin-1 (XPO-1) shuttles FXR1 to the cytoplasm, where RGG motifs facilitate binding to DNA or RNA structures. FXR1 is ubiquitously expressed and undergoes various post-translational modifications, including SUMOylation, ubiquitylation, acetylation, methylation, and phosphorylation at S420.<sup>10,11</sup>

FXR1 regulates microRNA (miRNA) processing, stabilizes specific miRNAs, and influences their subcellular localization.<sup>12</sup> It interacts with nucleoporins during mitosis, affecting protein export. Additionally, FXR1 stabilizes mRNA through adenylate–uridylylate (AU)-rich elements (AREs) in the 3′-untranslated region (3′-UTR), thereby influencing mRNA translation and stability. It also modulates mRNA translation through interactions with miRNAs, argonaute-2 (AGO2) and the RNA interference (RNAi) pathway. Furthermore, FXR1 has an influence on the cell cycle, synaptic protein function and muscle

development. It plays a crucial role in tumor necrosis factor alpha (TNF- $\alpha$ )–ARE-mediated translational activation, impacting cell growth. Overall, these multifaceted functions highlight the significance of FXR1 in post-transcriptional gene regulation and underscore its potential diagnostic and therapeutic implications.<sup>10,13</sup>

Recent studies have highlighted the involvement of FXR1 in cancer.<sup>14,15</sup> Elevated expression of FXR1 has been observed in various malignancies, including bladder cancer, colorectal cancer, non-small cell lung cancer, and hepatocellular carcinoma (HCC).<sup>14–18</sup> In these contexts, FXR1 acts as an oncogene, promoting tumor growth and invasion through mechanisms such as mRNA stabilization, enhanced protein translation and inactivation of tumor suppressor genes.<sup>19–21</sup>

Despite its pro-tumorigenic attributes, the RNA-binding mechanisms of FXR1 offer promising therapeutic avenues. Targeting these interactions using small molecules or RNAi-based strategies may disrupt its oncogenic effects. In HNSCC, genetic alterations, particularly copy number variation (CNV), play a crucial role in activating oncogenes or inactivating tumor suppressor genes.<sup>22–24</sup> Recent studies have also demonstrated the overexpression of FXR1 in HNSCC tissues, correlating it with tumor growth and invasion.<sup>25</sup> This study, utilizing data from The Cancer Genome Atlas (TCGA)–HNSCC cohort, aimed to analyze *FXR1* genetic variations, including the expression of DNA, mRNA, protein, as well as to unravel its molecular role and therapeutic potential in HNSCC. FXR1 is significantly associated with HNSCC, thus providing a link between the prognosis and tumor infiltration in HNSCC.

## Material and methods

### Copy number variation analysis

The cBioPortal (<https://www.cbioportal.org>), a user-friendly and interactive online resource, facilitates the visualization, analysis and download of extensive cancer genomics datasets.<sup>26</sup> In this investigation, the cBioPortal was used to examine alterations in the FXR1 gene among

HNSCC patients. Comprehensive clinical data for 528 HNSCC patients is presented in Table 1. The prognostic significance of *FXR1* genetic alterations in patients with HNSCC was assessed using the Kaplan–Meier plotter analysis.

## FXR1 expression analysis

The University of Alabama at Birmingham CANcer data analysis Portal (UALCAN) (<http://ualcan.path.uab.edu>)<sup>27</sup> was used to analyze expression profiles in HNSCC. The UALCAN integrates datasets from TCGA and the Clinical Proteomic Tumor Analysis Consortium (CPTAC).

**Table 1.** Demographic and clinical characteristics of The Cancer Genome Atlas (TCGA)–head and neck squamous cell carcinoma (HNSCC) cohort used for *FXR1* analysis ( $N = 528$ )

	Variable	Patients, $n$ (%)
Sex	male	383 (72.5)
	female	136 (25.8)
	not available	9 (1.7)
Age [years]	21–40	20 (3.8)
	41–60	236 (44.7)
	61–80	237 (44.9)
	81–100	24 (4.5)
	not available	11 (2.1)
Patient race	Caucasian	444 (84.1)
	African American	47 (8.9)
	Asian	11 (2.1)
	not available	26 (4.9)
Tumor grade	well differentiated (grade 1)	62 (11.7)
	moderately differentiated (grade 2)	303 (57.4)
	poorly differentiated (grade 3)	125 (23.7)
	undifferentiated (grade 4)	7 (1.3)
	not available	31 (5.9)
Cancer stage	1	27 (5.1)
	2	71 (13.4)
	3	81 (15.3)
	4	264 (50.0)
	not available	85 (16.1)
Lymph node metastasis status	no regional lymph node metastasis (N0)	176 (33.3)
	metastases in 1–3 axillary lymph nodes (N1)	67 (12.7)
	metastases in 4–9 axillary lymph nodes (N2)	12 (2.3)
	metastases in $\geq 10$ axillary lymph nodes (N3)	8 (1.5)
	not available	265 (50.2)
HPV status	positive	80 (15.2)
	negative	434 (82.2)
	not available	14 (2.7)
TP53 mutation status	TP53 mutant	327 (61.9)
	TP53 non-mutant	175 (33.1)
	not available	26 (4.9)

HPV – human papillomavirus.

The present study also involved a comprehensive analysis of the relationship between *FXR1* expression and clinicopathological features of HNSCC, such as disease stage, histological grade, lymph node involvement, and HPV status.

## Immunohistochemistry analysis

The Human Protein Atlas (HPA) (<http://www.proteinatlas.org>)<sup>28</sup> is a database that provides information on human protein-coding genes, including RNA and protein expression levels as well as location. Immunohistochemistry (IHC) images illustrating the expression of *FXR1* protein in clinical specimens from HNSCC patients and normal tissues were obtained from the HPA database. To analyze the expression levels of *FXR1*, staining intensity was classified as low, moderate or high. According to HPA data, Anti-*FXR1* rabbit monoclonal antibodies (HPA018246 and HPA055475; Sigma-Aldrich, St. Louis, USA) were utilized for IHC, with dilution ratios of 1:150 and 1:50, respectively.

## Immune infiltration analysis

Given the pivotal role of immune genes in the pathogenesis of HNSCC, we analyzed the relationship between our target gene, *FXR1*, and key immune genes involved in HNSCC pathogenesis. Tumor infiltration was assessed using TIMER 2.0 (<http://TIMER.cistrome.org>)<sup>29</sup> and the TISIDB (<http://cis.hku.hk/TISIDB>)<sup>30</sup> database. The TIMER server was employed to systematically evaluate immune infiltration across various malignancies, employing the TIMER method to quantify CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells, B cells, neutrophils, macrophages, and dendritic cells. The gene module was applied to examine *FXR1* expression in different cancers and the extent of immune cell infiltration in HNSCC. Additional analyses included the exploration of immunomodulators, chemokines and their receptors. Specifically, the top 3 negatively correlated immunomodulatory genes, encompassing major histocompatibility complex (MHC) molecules, immunostimulators and immune inhibitors, were identified in relation to *FXR1* mRNA expression in HNSCC patients.

## Statistical analysis

Data was analyzed using various statistical methods depending on the specific research question. To compare *FXR1* expression between the groups, Student's *t*-tests were performed to calculate *p*-values and determine fold changes, along with gene rank information. The Kaplan–Meier analysis was used to assess the relationship between variables and survival outcomes, with the log-rank test applied for comparisons between survival curves. To comprehensively evaluate the impact of *FXR1* expression and other clinicopathological factors on survival, a multivariate Cox proportional hazards model was employed.

Correlations between gene expression levels were assessed using Spearman's correlation analysis in the TIMER 2.0 databases. The TISIDB database provided both Spearman's and Pearson's correlation analyses to evaluate the strength of association between variables. A  $p$ -value  $<0.05$  was considered statistically significant unless stated otherwise.

## Results

### Association between genetic alterations in *FXR1* and increased mRNA expression in HNSCC

Genetic alterations in *FXR1* were observed in 22% of HNSCC patients among the 528 individuals analyzed. These alterations were primarily characterized by *FXR1* gene amplification, while some cases exhibited missense mutations (Fig. 1A). Notably, *FXR1* amplification was significantly correlated with elevated *FXR1* mRNA expression in HNSCC patients (Fig. 1B) ( $p < 0.05$ ).

### Upregulation of *FXR1* mRNA and protein levels in HNSCC

The mRNA expression level of *FXR1* was significantly upregulated in patients with primary HNSCC tumors ( $n = 520$ ) compared to normal tissues ( $n = 44$ ) in the

TCGA–HNSCC dataset (Fig. 2A) ( $p < 0.001$ ). Similarly, *FXR1* protein levels were significantly increased in primary HNSCC tumor tissues ( $n = 108$ ) compared to normal tissues ( $n = 71$ ) in the CPTAC–HNSCC dataset (Fig. 2B) ( $p < 0.001$ ). Immunohistochemical analysis further confirmed that *FXR1* protein was markedly higher in HNSCC tumor tissues than in normal tissues (Fig. 2C). Moreover, the Kaplan–Meier curve displayed the relationship between *FXR1* genetic alterations and HNSCC patient survival. In particular, patients with altered *FXR1* exhibited poorer overall survival, suggesting its potential prognostic significance (Fig. 2D) ( $p < 0.05$ ).

### Association of dysregulated *FXR1* mRNA with stage, grade, metastasis status, and HPV status in HNSCC

*FXR1* mRNA expression was significantly altered across different tumor stages, grades and metastasis statuses compared to normal tissues (Fig. 3A–C) ( $p < 0.05$ ). In terms of tumor stage, significant differences were noted between normal tissues and stages 2, 3 and 4, as well as between stages 3 and 4 (Fig. 3A) ( $p < 0.05$ ). Regarding HNSCC grade, substantial alterations were observed between normal tissues and grades 2, 3 and 4, as well as between grade 1 and grades 2 and 3 (Fig. 3B) ( $p < 0.05$ ). In terms of metastasis status, *FXR1* expression in normal tissues differed significantly from N0–N3 groups.

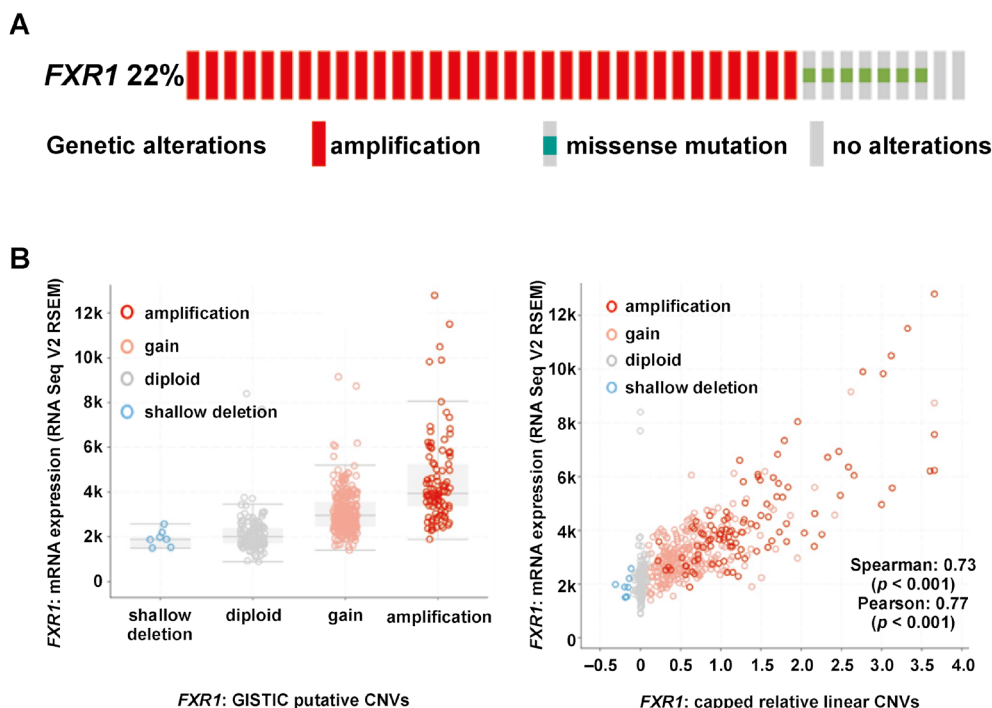
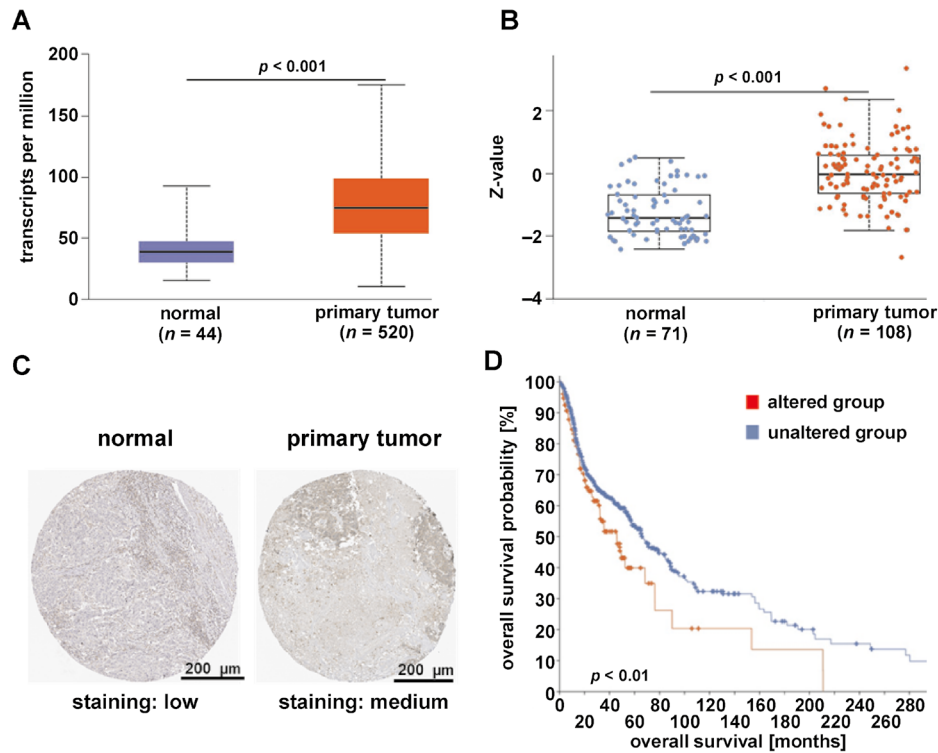


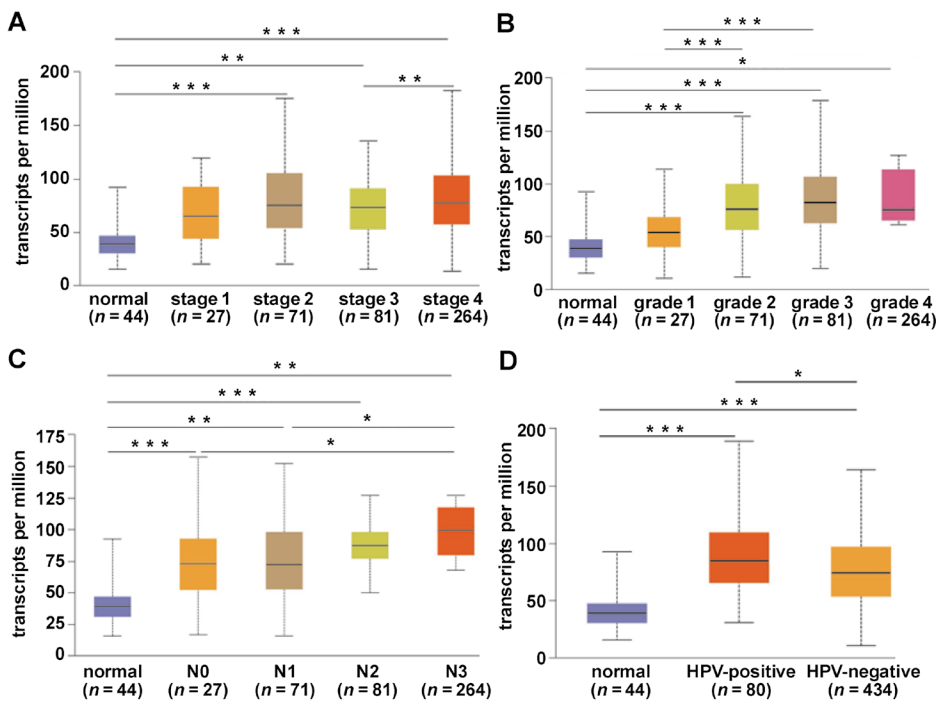
Fig. 1. *FXR1* gene alterations in head and neck squamous cell carcinoma (HNSCC) (source: cBioPortal)

A. OncoPrint illustrating genomic alterations of *FXR1* in HNSCC patients; B. Correlation between *FXR1* mRNA expression and copy number variations (CNVs) in HNSCC tissues, assessed using Spearman's and Pearson's correlation analyses. RSEM – RNA-Seq by Expectation Maximization.



**Fig. 2.** FXR1 expression and prognosis in head and neck squamous cell carcinoma (HNSCC) (sources: University of Alabama at Birmingham CANCER data analysis Portal (UALCAN), Human Protein Atlas (HPA) and cBioPortal)

A. *FXR1* mRNA expression in HNSCC tumor tissues compared to normal tissues (The Cancer Genome Atlas (TCGA)–HNSCC cohort); B. FXR1 protein expression in HNSCC tumor tissues compared to normal tissues (Clinical Proteomic Tumor Analysis Consortium (CPTAC)–HNSCC cohort); C. Immunohistochemical staining confirming elevated FXR1 protein expression in HNSCC tumor tissues compared to normal tissues (TCGA–HNSCC cohort); D. Overall survival of patients with *FXR1* genetic alterations compared to those without alterations (TCGA–HNSCC cohort).



**Fig. 3.** *FXR1* expression and clinicopathological features in head and neck squamous cell carcinoma (HNSCC) (source: University of Alabama at Birmingham CANCER data analysis Portal (UALCAN))

A. *FXR1* mRNA expression according to cancer stage; B. *FXR1* mRNA expression according to tumor grade; C. *FXR1* mRNA expression according to lymph node metastasis status; D. *FXR1* mRNA expression in human papillomavirus (HPV)-positive and HPV-negative HNSCC patients.

\*  $p < 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

Additionally, significant differences were observed between N0 and N3, and between N1 and N3 (Fig. 3C) ( $p < 0.05$ ). Furthermore, *FXR1* expression was significantly higher in HPV-positive HNSCC patients compared to HPV-negative patients and normal tissues (Fig. 3D) ( $p < 0.05$ ).

### Impact of *FXR1* mRNA expression on tumor immune infiltration in HNSCC

The mRNA expression of *FXR1* exhibited a significant negative correlation with CD8<sup>+</sup> T cells (Fig. 4A). High *FXR1* expression was associated with poorer prognosis in HNSCC patients, including those with HPV-negative tumors (Fig. 4A,B). The mRNA expression of *FXR1* and genes involved in MHC molecules, immunostimulators and immunoinhibitors, were analyzed in HNSCC patients, as depicted in Supplementary Figure 1 (available on request from the corresponding author). These groups function as immunomodulators in humans. The top 3 negatively correlated genes in each category were identified, with MHC molecules (*HLA-DMA*, *HLA-DPB1*, *HLA-DRB1*), immunostimulators (*KLRK1*, *CD27*, *TNFSF13B*) and immune inhibitors (*CD244*, *CTLA4*, *CD96*) showing significant negative correlations with *FXR1* expression in HNSCC patients (Fig. 5A–C) ( $p < 0.001$ ).

## Discussion

Among RNA-binding proteins, FXR1 is a well-characterized oncogene involved in multiple cancers. The overexpression of FXR1 has been reported in several malignancies, including glioma, lung carcinoma, HCC, breast cancer, head and neck carcinoma, ovarian cancer, and colorectal carcinoma.<sup>14,15,17,18,20,25</sup>

Copy number variation has been associated with changes in gene expression; yet, the specific cellular associations between CNV and gene expression alterations remain unknown.<sup>31</sup> The present study investigated genetic alterations and their functional implications, as well as the association between *FXR1* mRNA expression and HNSCC. Genetic alterations in *FXR1* were observed in 22% of HNSCC patients and were primarily characterized by gene amplification, with a smaller proportion of cases exhibiting missense mutations. Notably, *FXR1* amplification was significantly correlated with elevated *FXR1* mRNA expression, indicating a potential regulatory mechanism. This observation confirms previous findings demonstrating the influence of gene copy number on expression levels. Additionally, earlier studies have reported enrichment of both amplified and downregulated genes within the ubiquitin–proteasome system, suggesting potential consequences for protein degradation pathways in HNSCC.<sup>31</sup> Further research by Baltaci et al.<sup>32</sup> and Carron et al.<sup>33</sup> demonstrated that

CNVs, particularly gene amplification, contribute to HNSCC carcinogenesis through the cooperation of oncogenes and inherited CNV, influencing the occurrence and prognosis of oropharyngeal cancer, respectively. Beyond oncology, genetic alterations in *FXR1* have also been implicated in congenital multiminicore myopathy and Fragile X syndrome.<sup>34,35</sup>

FXR1 emerges as a potential biomarker and immune regulator in HNSCC. Both mRNA and protein levels were elevated in tumor tissues compared to healthy tissues, suggesting a role in disease progression. This upregulation is associated with patient survival and correlates with diverse clinical features, including HPV status. Previous studies have examined the role of FXR1 in cancer. Xiao et al. demonstrated the importance of *FXR1* across 25 cancer types, including breast and cervical cancers, showcasing its diagnostic potential and association with prognosis.<sup>36</sup> The enrichment analysis revealed the influence of FXR1 on tumor development and progression through multiple pathways such as cell cycle control, mitochondrial metabolism, immune response, and skin barrier formation.<sup>36</sup> Notably, RBPs have emerged as key regulators of these processes, influencing RNA splicing, transport, translation, and localization.<sup>37</sup> Further studies have demonstrated the diverse roles of FXR1, ranging from influencing RNA metabolism in glioblastoma to impacting muscle development and adenovirus interactions.<sup>20,38,39</sup> These findings reveal the broad involvement of FXR1 in various cancer types and disease-related processes.

While existing research explores the role of FXR1 in cancer, several key mechanisms remain shrouded. In ovarian cancer, FXR1 has been shown to regulate c-MYC translation and stabilization, promoting its oncogenic activity.<sup>40</sup> Moreover, cancer cells lacking both p53 and FXR2 (“double knockout”) exhibit increased sensitivity to FXR1 inhibition. This vulnerability depends on the FXR1 tandem Tudor domain and its interaction with STAT1/3 proteins. FXR1 binds to gene promoters, including *ARID1A* and *CASC4*, and recruits STAT1/3 to promote transcription. The inhibition of FXR1 or JAK (activating STAT1/3) suppresses FXR1 target genes and reduces cancer cell growth.<sup>41</sup> These findings reveal a novel synergy between FXR1 and STAT1/3 in regulating cell proliferation, opening therapeutic avenues for double-knockout cancers. Furthermore, in HCC, FXR1 has been implicated in the intricate transforming growth factor beta (TGF- $\beta$ )/SMAD signaling pathway. Elevated FXR1 expression correlates with dismal prognosis and promotes cancer cell motility, invasion and proliferation through the modulation of the SMAD2/3 axis.<sup>18</sup> Moreover, Majumder et al. highlighted the ability of FXR1 to bypass cellular senescence in oral cancer, revealing its contribution to tumor cell survival and growth.<sup>42</sup> These outcomes suggest that FXR1 presents additional therapeutic opportunities across diverse cancer landscapes.

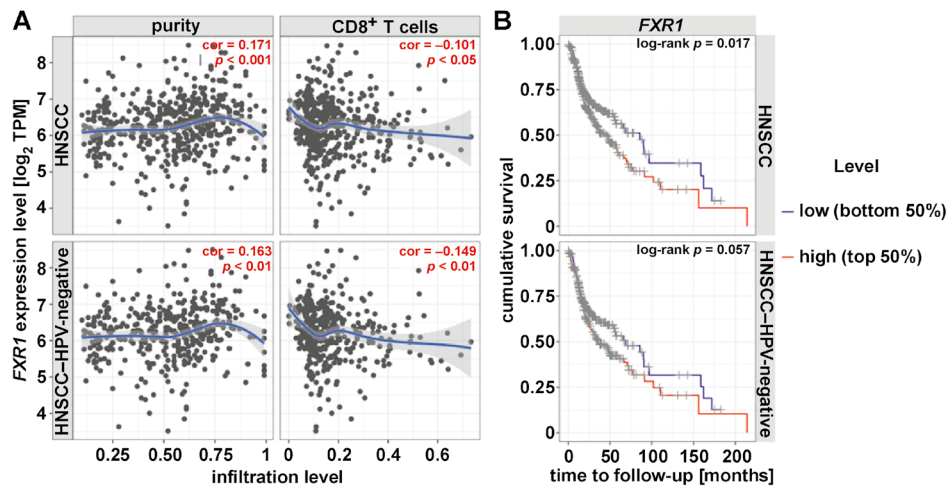


Fig. 4. *FXR1* expression and immune infiltration in head and neck squamous cell carcinoma (HNSCC) (source: TIMER 2.0)

A. Correlation between *FXR1* expression and CD8<sup>+</sup> T cell infiltration in HNSCC patients; B. Association between *FXR1* expression and survival outcomes in HNSCC patients.

TPM – transcripts per million.

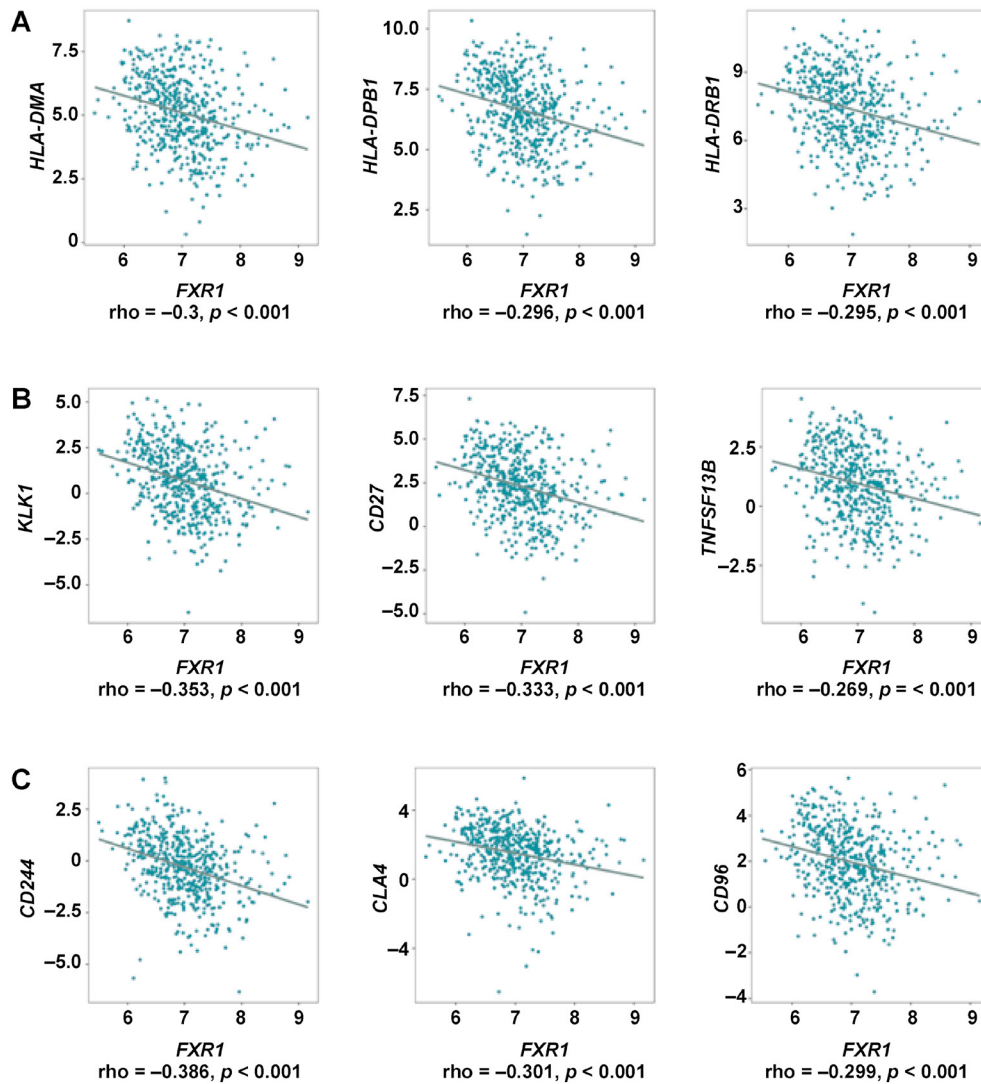


Fig. 5. Expression of immune regulatory genes and *FXR1* in head and neck squamous cell carcinoma (HNSCC) (source: TISIDB; Spearman’s correlation analysis,  $n = 522$ )

A. Major histocompatibility complex (MHC) molecule genes; B. Immunostimulatory genes; C. Immune inhibitory genes.

A growing number of studies have shown that the immune microenvironment plays a vital role in the development and progression of HNSCC.<sup>43</sup> Previous studies have shown that *FXR1* expression is significantly correlated with the expression of immunomodulators across various cancers.<sup>36</sup> In this study, *FXR1* expression was negatively correlated with CD8<sup>+</sup> T cells, implying a role in the modulation of the immune microenvironment. Further analyses indicated that *FXR1* interacts with key immune-related genes, supporting its potential as an immunotherapeutic target. These findings warrant further investigation into the multifaceted role of FXR1 in HNSCC progression and immune evasion.

The current study, based on the TCGA–HNSCC dataset and various bioinformatics tools, provides valuable insights into the potential of FXR1 as a biomarker and prognostic indicator in HNSCC. However, further research is required to fully grasp its potential. Large-scale cohort analysis of genetic variants and expression patterns across diverse experimental methods could significantly enhance the accuracy and robustness of these findings. Functional experiments and in vivo studies are crucial for validating the therapeutic potential of FXR1 in HNSCC. Our initial data has shed light on how genetic variants focusing on CNV influence the pathogenesis of HNSCC by altering FXR1 expression and regulation. Building upon this foundation, further research can translate these findings into tangible therapeutic strategies.

## Conclusions

This study demonstrates a significant association between *FXR1* genetic alterations and aberrant expression in HNSCC. *FXR1* gene amplification contributes to elevated FXR1 mRNA and protein levels in HNSCC patients. This upregulation is associated with poor prognosis and correlates with various clinical features, including disease stage, grade, lymph node metastasis status, and HPV status. Additionally, our findings suggest that FXR1 may play a role in modulating the immune microenvironment, emphasizing its multifaceted involvement in HNSCC progression and immune evasion. Taken together, FXR1 emerges as a promising biomarker and therapeutic target, warranting further exploration of its molecular mechanisms and therapeutic potential in HNSCC.

## Ethics approval and consent to participate

Not applicable.

## Data availability

The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Consent for publication


Not applicable.

## Use of AI and AI-assisted technologies


ChatGPT (OpenAI, San Francisco, USA) was used to improve readability. The content was then subjected to a thorough review and editing process to ensure the highest standards of quality. The authors accept full responsibility for the publication.


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## References

- Johnson DE, Burtneß B, Leemans CR, Lui VWY, Bauman JE, Grandis JR. Head and neck squamous cell carcinoma. *Nat Rev Dis Primers*. 2020;6(1):92. doi:10.1038/s41572-020-00224-3
- Sung H, Ferlay J, Siegel RL, et al. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2021;71(3):209–249. doi:10.3322/caac.21660
- Barsouk A, Aluru JS, Rawla P, Saginala K, Barsouk A. Epidemiology, risk factors, and prevention of head and neck squamous cell carcinoma. *Med Sci (Basel)*. 2023;11(2):42. doi:10.3390/medsci11020042
- Kannan B, Pandi C, Pandi A, Jayaseelan VP, Murugan MS, Arumugam P. Altered expression of GLS2 indicates a poor prognosis and correlates with clinicopathological features of oral squamous cell carcinoma. *Int J Oral Maxillofac Surg*. 2024;53(8):635–643. doi:10.1016/j.ijom.2024.01.011
- Avs KR, Pandi C, Kannan B, Pandi A, Jayaseelan VP, Arumugam P. RFC3 serves as a novel prognostic biomarker and target for head and neck squamous cell carcinoma. *Clin Oral Investig*. 2023;27(11):6961–6969. doi:10.1007/s00784-023-05316-4
- Balachander K, Paramasivam A. Ferroptosis: An emerging therapeutic target for oral cancer. *Oral Oncol*. 2022;131:105970. doi:10.1016/j.oraloncology.2022.105970
- Romanowska K, Sobiecka A, Rawluszko-Wieczorek AA, Suchowska WM, Golusiński W. Head and neck squamous cell carcinoma: Epigenetic landscape. *Diagnostics (Basel)*. 2020;11(1):34. doi:10.3390/diagnostics11010034
- Ju Y, Wu X, Wang H, et al. Genomic landscape of head and neck squamous cell carcinoma across different anatomic sites in Chinese population. *Front Genet*. 2021;12:680699. doi:10.3389/fgene.2021.680699
- Siomi MC, Siomi H, Sauer WH, Srinivasan S, Nussbaum RL, Dreyfuss G. FXR1, an autosomal homolog of the fragile X mental retardation gene. *EMBO J*. 1995;14(11):2401–2408. doi:10.1002/j.1460-2075.1995.tb07237.x
- Khan FA, Fang N, Zhang W, Ji S. The multifaceted role of Fragile X-Related Protein 1 (FXR1) in cellular processes: An updated review on cancer and clinical applications. *Cell Death Dis*. 2024;15(1):72. doi:10.1038/s41419-023-06413-8
- Majumder M, Johnson RH, Palanisamy V. Fragile X-related protein family: A double-edged sword in neurodevelopmental disorders and cancer. *Crit Rev Biochem Mol Biol*. 2020;55(5):409–424. doi:10.1080/10409238.2020.1810621
- van Kouwenhove M, Kedde M, Agami R. MicroRNA regulation by RNA-binding proteins and its implications for cancer. *Nat Rev Cancer*. 2011;11(9):644–656. doi:10.1038/nrc3107
- St Paul A, Corbett C, Peluzzo A, et al. FXR1 regulates vascular smooth muscle cell cytoskeleton, VSMC contractility, and blood pressure by multiple mechanisms. *Cell Rep*. 2023;42(4):112381. doi:10.1016/j.celrep.2023.112381

14. Qian J, Hassanein M, Hoeksema MD, et al. The RNA binding protein FXR1 is a new driver in the 3q26-29 amplicon and predicts poor prognosis in human cancers. *Proc Natl Acad Sci U S A*. 2015;112(11):3469–3474. doi:10.1073/pnas.1421975112
15. Comtesse N, Keller A, Diesinger I, et al. Frequent overexpression of the genes *FXR1*, *CLAPM1* and *EIF4G* located on amplicon 3q26-27 in squamous cell carcinoma of the lung. *Int J Cancer*. 2007;120(12):2538–2544. doi:10.1002/ijc.22585
16. Deng M, Wang N, Li Z, et al. FXR1 can bind with the CFIm25/CFIm68 complex and promote the progression of urothelial carcinoma of the bladder by stabilizing TRAF1 mRNA. *Cell Death Dis*. 2022;13(2):170. doi:10.1038/s41419-022-04614-1
17. Jin X, Zhai B, Fang T, Guo X, Xu L. FXR1 is elevated in colorectal cancer and acts as an oncogene. *Tumour Biol*. 2016;37(2):2683–2690. doi:10.1007/s13277-015-4068-9
18. Zhao K, Gao J, Shi J, et al. FXR1 promotes proliferation, invasion and migration of hepatocellular carcinoma in vitro and in vivo. *Oncol Lett*. 2023;25(1):22. doi:10.3892/ol.2022.13608
19. Cao H, Gao R, Yu C, Chen L, Feng Y. The RNA-binding protein FXR1 modulates prostate cancer progression by regulating FBXO4. *Funct Integr Genomics*. 2019;19(3):487–496. doi:10.1007/s10142-019-00661-8
20. Cao S, Zheng J, Liu X, et al. FXR1 promotes the malignant biological behavior of glioma cells via stabilizing MIR17HG. *J Exp Clin Cancer Res*. 2019;38(1):37. doi:10.1186/s13046-018-0991-0
21. Datta C, Truesdell SS, Wu KQ, et al. Ribosome changes reprogram translation for chemosurvival in G0 leukemic cells. *Sci Adv*. 2022;8(43):eabo1304. doi:10.1126/sciadv.abo1304
22. Yang J, Chen Y, Luo H, Cai H. The landscape of somatic copy number alterations in head and neck squamous cell carcinoma. *Front Oncol*. 2020;10:321. doi:10.3389/fonc.2020.00321
23. Chen Y, Chen C. DNA copy number variation and loss of heterozygosity in relation to recurrence of and survival from head and neck squamous cell carcinoma: A review. *Head Neck*. 2008;30(10):1361–1383. doi:10.1002/hed.20861
24. Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. *Arch Oral Biol*. 2021;122:105030. doi:10.1016/j.archoralbio.2020.105030
25. Qie S, Majumder M, Mackiewicz K, et al. Fbxo4-mediated degradation of Fxr1 suppresses tumorigenesis in head and neck squamous cell carcinoma. *Nat Commun*. 2017;8(1):1534. doi:10.1038/s41467-017-01199-8
26. de Bruijn I, Kundra R, Mastrogiacomo B, et al. Analysis and visualization of longitudinal genomic and clinical data from the AACR Project GENIE Biopharma collaborative in cBioPortal. *Cancer Res*. 2023;83(23):3861–3867. doi:10.1158/0008-5472.CAN-23-0816
27. Chandrashekar DS, Karthikeyan SK, Korla PK, et al. UALCAN: An update to the integrated cancer data analysis platform. *Neoplasia*. 2022;25:18–27. doi:10.1016/j.neo.2022.01.001
28. Uhlen M, Zhang C, Lee S, et al. A pathology atlas of the human cancer transcriptome. *Science*. 2017;357(6352):eaan2507. doi:10.1126/science.aan2507
29. Li T, Fu J, Zeng Z, et al. TIMER2.0 for analysis of tumor-infiltrating immune cells. *Nucleic Acids Res*. 2020;48(W1):W509–W514. doi:10.1093/nar/gkaa407
30. Ru B, Wong CN, Tong Y, et al. TISIDB: An integrated repository portal for tumor-immune system interactions. *Bioinformatics*. 2019;35(20):4200–4202. doi:10.1093/bioinformatics/btz210
31. Shao X, Lv N, Liao J, et al. Copy number variation is highly correlated with differential gene expression: A pan-cancer study. *BMC Med Genet*. 2019;20(1):175. doi:10.1186/s12881-019-0909-5
32. Baltaci E, Karaman E, Dalay N, Buyru N. Analysis of gene copy number changes in head and neck cancer. *Clin Otolaryngol*. 2018;43(4):1004–1009. doi:10.1111/coa.12686
33. Carron J, Torricelli C, Silva JK, et al. Association of inherited copy number variation in *ADAM3A* and *ADAM5* pseudogenes with oropharynx cancer risk and outcome. *Genes (Basel)*. 2022;13(12):2408. doi:10.3390/genes13122408
34. Estañ MC, Fernández-Núñez E, Zaki MS, et al. Recessive mutations in muscle-specific isoforms of FXR1 cause congenital multi-minicore myopathy. *Nat Commun*. 2019;10(1):797. doi:10.1038/s41467-019-08548-9
35. Zang JB, Nosyreva ED, Spencer CM, et al. A mouse model of the human Fragile X syndrome I304N mutation. *PLoS Genet*. 2009;5(12):e1000758. doi:10.1371/journal.pgen.1000758
36. Xiao K, Ullah I, Yang F, et al. Comprehensive bioinformatics analysis of FXR1 across pan-cancer: Unraveling its diagnostic, prognostic, and immunological significance. *Medicine (Baltimore)*. 2023;102(48):e36456. doi:10.1097/MD.00000000000036456
37. Mangus DA, Evans MC, Jacobson A. Poly(A)-binding proteins: Multifunctional scaffolds for the post-transcriptional control of gene expression. *Genome Biol*. 2003;4(7):223. doi:10.1186/gb-2003-4-7-223
38. Smith JA, Curry EG, Blue RE, et al. FXR1 splicing is important for muscle development and biomolecular condensates in muscle cells. *J Cell Biol*. 2020;219(4):e201911129. doi:10.1083/jcb.201911129
39. Hajikhezri Z, Kaira Y, Schubert E, et al. Fragile X-related protein FXR1 controls human adenovirus capsid mRNA metabolism. *J Virol*. 2023;97(2):e0153922. doi:10.1128/jvi.01539-22
40. George J, Li Y, Kadamberi IP, et al. RNA-binding protein FXR1 drives cMYC translation by recruiting eIF4F complex to the translation start site. *Cell Rep*. 2021;37(5):109934. doi:10.1016/j.celrep.2021.109934
41. Fan Y, Yue J, Xiao M, et al. FXR1 regulates transcription and is required for growth of human cancer cells with *TP53/FXR2* homozygous deletion. *Elife*. 2017;6:e26129. doi:10.7554/eLife.26129
42. Majumder M, House R, Palanisamy N, et al. Correction: RNA-binding protein FXR1 regulates p21 and TERC RNA to bypass p53-mediated cellular senescence in OSCC. *PLoS Genet*. 2016;12(10):e1006411. doi:10.1371/journal.pgen.1006411
43. Chen SMY, Krinsky AL, Woolaver RA, Wang X, Chen Z, Wang JH. Tumor immune microenvironment in head and neck cancers. *Mol Carcinog*. 2020;59(7):766–774. doi:10.1002/mc.23162